EXact-Cut™ Mlul Restriction Endonuclease

Catalog Number: EXNA022

Size: 1,000 Units

For Research Use Only. Not Intended for Diagnostic or Therapeutic Use.



Product Details

Description	EXact-Cut [™] Mlul Restriction Endonuclease is engineered for high specificity, reduced star activity, and time saving DNA digestion in 5-15 minutes. To simplify experimental design using multiple restriction enzymes, our entire range of EXact-Cut [™] restriction endonucleases are 100% active in our EXact-Cut [™] buffer (included) and are optimized for single-tube reactions along with digestion and ligation protocols.			
Restriction Enzyme Site	5'A↓CGCG T3' 3'T GCGC↑A5'			
Unit Definition	One unit is defined as the amount of enzyme required to digest 1 μg of λ DNA in 1 hour at 37°C in a total reaction volume of 50 μL .			
Recommended Reaction Conditions	1X EXact-Cut™ Buffer Incubate at 37°C Refer to Protocol for reaction setup			
Heat Inactivation	 Incubate at 80°C for 20 minutes Add appropriate volume of 6X Gel Loading Dye, according to the reaction system 			
Components	EXact-Cut™ MluI (10 Units/μL) EXact-Cut™ 10X Buffer 6X Gel Loading Dye, Purple	1,000 Units 1 mL 1 mL		

Storage and Preparation

Shipping	Shipped on blue ice.
Stability and Storage	Store at -20°C for up to 24 months.

Protocol

Protocol for Rapid DNA Digestion

1. Add the following components on ice in the indicated order:

	Plasmid DNA	PCR Product	Genomic DNA
DNA	≤ 1 µg	≤ 0.2 µg	≤ 5 µg
EXact-Cut™ 10X Buffer	2 μL	3 μL	5 μL
ddH ₂ O, make up to final volume indicated:	20 μL	30 μL	50 μL
Exact-Cut™ MluI	10 Units	10 Units	30-50 Units

Note: DNA should be free of phenol, chloroform, ethanol, EDTA, detergents or high concentrations of salts. For compatibility with other common buffers, see the chart on page 2.

- 2. Gently mix or flick the tube to mix (do not vortex), then immediately follow with a guick spin-down in a microcentrifuge.
- 3. Incubate at 37°C for the indicated sample type: plasmid DNA (15 minutes), PCR product (15-30 minutes), or genomic DNA (30-60 minutes)
- 4. Optional: inactivate the enzyme at 80°C for 20 minutes and add an appropriate amount of 6X Gel Loading Dye, according to the reaction system.

Protocol for Multiple Digestion of DNA

- 1. Use 10 Units of each enzyme and scale up to the reaction conditions accordingly.
- 2. The combined volume of the enzymes in the reaction mixture **should not** exceed **1/10** of the total reaction volume.
- 3. If the enzymes require different reaction temperatures, start with the enzyme requiring the lowest temperature, followed by the next enzyme(s) and incubate at the higher temperature.

Note: For total reaction volumes > 20 μL, the incubation time should be increased accordingly in a water bath.



2726 Summer Street NE

Number of Recognition Sites in DNA										
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λDNA	ФХ174	pBR322	pUC57	pUC18/19	SV40	M13mp18/19	Adeno2			
7	2	0	0	0	0	0	5			
Methylation	Methylation Effects on Digestion									
Dam		Dcm	CpG		EcoKI	oKI EcoBI				
No effec	No effect No effect Blocked		ocked	No effect	No	No effect				
Activity in Common Buffers										
		EXact-Cut™ Buffer		akara ut™ Buffer	Thermo Scientif		NEB art® Buffer			
Activity	,	100%	10	00%	75%	,	100%			
Application	Notes									
Functional Test A 20 μL reaction in EXact-Cut Buffer containing 1 μg of λDNA and 10 Units of EXact-Cut Mlul incubated for 15 minutes at 37°C results in complete digestion as determined by agarose gel electrophoresis.										
Digestion-Liga	tion Test	At the optimal reaction temperature, the DNA was digested using 10 Units of EXact-Cut Mlul and the digestion product was recovered. The DNA fragments were ligated using an appropriate amount of T4 DNA Ligase at 22°C. After the ligation product was recovered, it was able to be recut with EXact-Cut Mlul.								
Non-Specific Endonuclease Test	Activity	At the optimal reaction temperature,10 Units of EXact-Cut MluI was incubated in 20 μ L reaction volume in EXact-Cut Buffer with 1 μ g of supercoiled plasmid DNA for 4 hours. Undigested, supercoiled plasmid DNA was detected using agarose gel electrophoresis.								